



Chemoenzymatic preparation of 4-*O*-acetyl sialic acid derivative

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Abstract

4-*O*-Acetyl sialic acid derivative could be easily prepared from the hydrolysis of **1** by lipase OF via the migration of acetyl groups. © 2000 Elsevier Science Ltd. All rights reserved.

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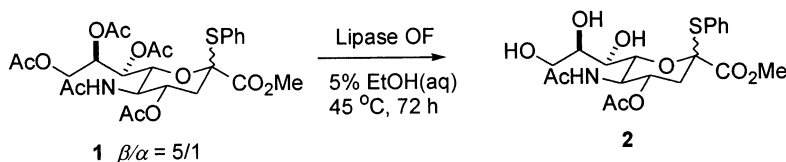
Sialic acids are the essential components of glycoconjugates which are important in cell–cell recognition at cell surfaces.¹ Many derivatives of sialic acids have been isolated and identified. Among them, the most common modification in sialic acids is the acetylation of the hydroxyl groups to form *O*-acetyl esters. More than 25 different acetylated sialic acids have been found in nature, and some have been demonstrated to involve biological functions.² For example, 4-*O*-acetylated sialic acids are resistant to all known sialidase,³ and 9-*O*-acetyl sialic acid in the non-reducing terminal glycoconjugates would generate a new epitope related to human melanoma cells.⁴ Since 4-*O*-acetylsialyl glycosides exhibit a remarkable inhibition of influenza A virus,⁵ the 4-*O*-acetyl sialyl derivative could be the key intermediates to develop anti-influenza drugs. In this communication, we developed a novel, simple and efficient chemoenzymatic method to prepare **2**, a 4-*O*-acetyl sialyl derivative.

Compound **1**, which serves as the starting material for the chemoenzymatic transformations, was prepared according to the procedure of Marra and Sinaÿ.⁶

A straightforward route to prepare compound **2** was obtained via direct deacetylation of **1** (a mixture of β/α diastereomers) using lipase OF (*Candida rugosa*) in a one-pot procedure (Scheme 1). After stirring a solution of **1** (119 mg, 0.204 mmol, $\beta/\alpha=5/1$) with lipase OF (0.3 g) in phosphate buffer solution (pH 7, 7 mL) containing 5% ethanol for 72 h at 45°C, compound **2** was isolated in 79% combined yield.⁷ According to our studies, lipase OF had the ability to remove acetyl groups in primary alcohol, but not in the secondary alcohol. Therefore, the acetyl

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group in C-9 of **1** was the first acetyl group to be removed. Based on an earlier report of acetyl migration in sialic acid,^{2,8} the acetyl groups at the C-8 and C-7 positions would then migrate to the C-9 position where they would be cleaved by the enzyme to generate **2**.⁹ The C-4 acetyl group was remarkably stable in this lipase-catalyzed hydrolysis.



Scheme 1.

In conclusion, it is noteworthy to emphasize that compared with the previous reports,¹⁰ this chemoenzymatic method can be used to efficiently prepare **2**, which is an important building block (donor) in the synthesis of 4-*O*-acetylsialyl glycosides.

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References

- (a) Lowe, J. B. In *Molecular Glycobiology*; Fukuda, M.; Hindsgaul, O., Eds.; IRL Press at Oxford University Press: Oxford, 1994; Chapter 4. (b) Varki, A. *Proc. Natl. Acad. Sci.* **1994**, *91*, 7390. (c) Feizi, T. *Nature* **1985**, *314*, 53. (d) Edemann, G. M. *Science* **1983**, *219*, 450.
- Varki, A. *Glycobiology* **1992**, *2*, 25.
- Hanaoka, K.; Pritchett, T. J.; Takasaki, S.; Kochibe, N.; Sabesan, S.; Paulson, J. C.; Kobata, A. *J. Biol. Chem.* **1989**, *264*, 9842.
- Cheresh, D. A.; Reisfeld, R. A.; Varki, A. *Science* **1984**, *225*, 844.
- Schreiner, E.; Zbiral, E.; Kleineidam, R. G.; Schauer, R. *Liebigs. Ann. Chem.* **1991**, 129, and references cited therein.
- Marra, A.; Sinaÿ, P. *Carbohydr. Res.* **1989**, *18*, 35.
- The two α/β isomers could be easily separated by a flash column chromatography eluted with a gradient of eluents ($\text{CH}_2\text{Cl}_2/\text{MeOH}$: 20/1 to 10/1). The α isomer is less polar relative to the β isomer. The α isomer (14% yield): a white solid. LRMS (EI) m/z 457 (M^+ , 55%), 348 (16%), 288 (7%). $[\alpha]_{\text{D}}^{26} -0.404$ (c 0.16, MeOH). $^1\text{H NMR}$ (400 MHz, D_2O) δ 7.38–7.54 (m, 5H), 4.88 (ddd, $J=13.1, 11.2, 5.0$ Hz, 1H), 4.09 (t, $J=10.5$ Hz, 1H), 3.70–3.79 (m, 2H), 3.65 (d, $J=10.6$ Hz, 1H), 3.59 (s, 3H), 3.50–3.57 (m, 2H), 2.91 (dd, $J=13.1, 5.0$ Hz, 1H), 2.02–2.07 (d+s, $J=12.6$ Hz, 4H), 1.90 (s, 3H). $^{13}\text{C NMR}$ (100 MHz, D_2O) δ 174.2, 172.9, 169.8, 136.2, 130.4, 128.8, 127.2, 86.3, 74.4, 71.0, 69.8, 67.5, 62.3, 53.0, 48.6, 36.2, 21.5, 19.8. The β isomer (65% yield): a white solid. HRMS (FAB) calcd for $\text{C}_{20}\text{H}_{27}\text{NO}_9\text{S}$ ($\text{M}+\text{Na}^+$) 480.1304, found 480.1298. $[\alpha]_{\text{D}}^{26} -138.14$ (c 0.255, MeOH). $^1\text{H NMR}$ (400 MHz, D_2O) δ 7.35–7.52 (m, 5H), 5.33 (ddd, $J=14.8, 11.3, 4.7$ Hz, 1H), 4.60 (d, $J=10.5$ Hz, 1H), 4.11 (t, $J=10.3$ Hz, 1H), 3.74–3.80 (m, 2H), 3.59–3.68 (m, 2H), 3.57 (s, 3H), 2.71 (dd, $J=13.7, 4.7$ Hz, 1H), 2.19 (dd, $J=13.7, 11.3$ Hz, 1H), 2.03 (s, 3H), 1.98 (s, 3H). $^{13}\text{C NMR}$ (100 MHz, D_2O) δ 174.2, 172.9, 169.8, 135.3, 129.8, 128.9, 128.2, 89.1, 70.9, 69.5, 67.9, 62.6, 52.9, 49.1, 36.2, 21.6, 19.9.
- Kamerling, J. P.; Schauer, R.; Shukla, A. K.; Stoll, S.; Van Beek, H.; Vliegthart, J. F. G. *Eur. J. Biochem.* **1987**, *162*, 601.

9. One of the examples of lipase catalyzed de-*O*-acetylation via migration of acetyl groups has been previously reported in this laboratory. See: Hsiao, K.-F.; Lin, H.-J.; Leu, D.-L.; Wu, S.-H.; Wang, K.-T. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 1629 and references 1–4 cited therein. The investigation of a variety of lipases in the deacetylation on different kinds of disaccharides is under the progress in this laboratory.
10. Shiba, T.; Yamamoto, T.; Teshima, T. In *Sialobiology and Other Novel Forms of Glycoylation*; Inoue, Y.; Lee, Y. C.; Troy II., F. A., Eds.; Gagushin, Osaka, 1999; pp. 289–301.